

Environmental Assessment and Planning of a Transboundary Fluvio-Marine Ecosystem: the Río de la Plata and its Maritime Front

CHAPTER 9. Fish Diversity of the Río de la Plata and its Maritime Front

FISHES FROM THE RÍO DE LA PLATA, THE ESTUARY AND ADJACENT SHELF AND SLOPE

Mirta L. García⁽¹⁾ Andrés J. Jaureguizar⁽²⁾, Hugo L. López⁽¹⁾, Amalia M. Miquelarena⁽¹⁾, Roberto C. Menni⁽¹⁾, Lucila C. Protogino⁽³⁾ and Graciela García⁽⁴⁾

⁽¹⁾ Museo de la Plata, Paseo del Bosque S/N. 1900. La Plata. Argentina.

⁽²⁾ Instituto Nacional de Investigación y Desarrollo Pesquero

⁽³⁾ Instituto de Limnología "Dr. Raúl Ringuelet"

[FALTA 4](#)

Email: mlgarcia@fcnym.unlp.edu.ar

ABSTRACT

The Río de La Plata has been traditionally divided in three zones, internal, intermediate and external, with clear geographic limits and distinctive environmental traits. The ichthyofauna is composed by freshwater, brackish and marine species which occurs along a longitudinal ecological gradient. Distribution of species throughout this gradient allows us to recognize two defined areas: a Riverine area characterized by a freshwater fish assemblage and an Estuarine area with an assemblage composed by freshwater, brackish water and marine fishes, including migrant species which use this space as nursery area. Following the gradient to the southwest we establish other two areas; one adjacent to the Río de la Plata (the Shelf area) and a following one (the Slope area), both with more

stable environmental and only marine fishes. Temperature and bottom salinity were identified as the determinant factors of these associations. Along the gradient, changes in the food energy source from benthic production or detritus to phytoplankton and zooplankton evidence their importance in structuring demersal fish communities. Analysis of population genetic structure analyses of commercial important fishes shows the existence of different genetic stocks which may be considered as management units in sustainable fisheries.

1. The fish fauna

It is somewhat peculiar that the fish taxonomy of the largest freshwater and brackish environment common to Argentina and Uruguay has not been really methodically studied. Probably the most important motive for this is its dimensions that except in coastal areas require large scale sampling methods. To this is added the lack of continuity and support to the creation of hydrobiological stations, in spite of proposal to that end since over a century (Popovici and Righi 1948). The best known part, which has been the subject of systematic research with oceanic methodologies, is the external zone (also marine section and/or estuarial zone). So, in lists of the Río de la Plata fishes, there are reported marine species which in fact strictly belong to brackish waters. The typical cases are lists that include only a few freshwater fishes (Marelli 1924, De Buen 1950, Cousseau 1985, Lasta 1995). Among the first fish reports for the Río de la Plata, are *Pimelodus maculatus* by Lacépède (1803), *Genidens barbatus* by Quoy and Gaimard in 1824, and *Odontesthes bonariensis*, *Loricariichthys anus* and *Hypostomus commersoni* by Valenciennes in 1835 and 1836 (Ringuelet et al. 1967).

Detailed lists of the Río de la Plata freshwater fishes were only available with the publication of lists by López et al. (1996) and Nion (1998), though many adequate data may be obtained from the book by Ringuelet et al. (1967), where the Río de la Plata is included as a single locality among many others. Many other localities in the river are reported by Menni et al. (1984). Any analysis of the available lists will clearly show that the fish fauna from the Río de la Plata is typically Brazilian or Paranensean in the sense of Ringuelet et al. (1967) and Ringuelet (1975), and that is very similar to that from the Paraná River.

The Río de la Plata, together with the Paraná River and the lower section of the Paraguay River, conform the Subtropical Potamic Axis (López et al. 2002) or the Great Rivers Province (López et al. 2008). The Paraná River Delta, with a well known Paranensean abundant fish fauna (164 species) (Liotta et al. 1995/1996), is adequate for comparative purposes. Nion (1998) listed 153 species from the Río de la Plata, 9 less than the Delta and Menni (2004) lists 166. In this paper we listed 161 species (Table 1) Environments of the Province of Buenos Aires (Argentina), connected with the Río de la Plata (Almirón et al. 2000) have a number of fish species about 30% of that from the River, a similar relation to that found between the Paraná River and related environments.

Considering that these environments in the Province of Buenos Aires, largely poorer than the Río de la Plata, have given origin to a enormous amount of empirical and theoretical information (Ringuelet 1962, 1975, references in López et al. 2006), we may have an idea of the magnitude of investigation required for an adequate knowledge of the Río de la Plata.

From a taxonomic point of view, the fish composition of the Río de la Plata has been naturally influenced by the important nomenclatural changes raised in the last decades, especially in the Characiformes. These changes may be followed through the lists published in different moments (Marelli 1924, Pozzi 1945, Ringuelet and Arámburu 1957, Ringuelet et al. 1967, Almirón et al. 1992, López et al. 1996, Nión 1998, Menni 2004).

The low variation in number of families and species of Siluriformes (except in relation to the oldest lists), show that the knowledge of species of this group is more complete than that of the other orders. In this case, as in others, it is possible that smaller or rarer species has been yet overlooked. Species number of Gymnotiformes practically doubled between first and latest lists. In other orders changes were lesser.

The number of species occurring in the Río de la Plata is about 35% of the total number given for Argentina (López et al. 2003). Among exotic fish, the carp is the most important because the wide distribution the species reached in few more than a half century (McDonagh 1945), invading inner waters of the Buenos Aires Province and totally occupying the Salado River Basin (López et al. 1996). More recent introductions were *A.cf. baerii* and *H. molitrix* (Azpelicueta and Almirón 1999, García Romero et al. 1998).

Cousseau (1985) provides a list of fishes mainly from the maritime front, including some species from the river itself, based in De Buen (1950), Abella et al. (1979), Ringuelet et al. (1967) and research trips from the Instituto Nacional de Investigación y Desarrollo Pesquero. She mentions six freshwater species (four siluriforms and two characiforms). García et al. (2003) report 19 chondrichthyan

species (all marine ones with the exception of *P. brachyura*), 170 freshwater species and 73 marine species.

To the West of the external zone there is large environment, the Samborombón Bay with a dense net of tide influenced environments which give to the bay physical, chemical and environmental traits that provides the basic conditions for the establishment of juveniles (Lasta 1995).

The Samborombón Bay includes retention, high productivity and nursering zones (Acha and Mianzan 2003). Lasta (1995) reports for bay five freshwater species (four siluriforms and a single caraciform). These numbers show that the ichthyofauna from the area is a combination of freshwater, brackish and marine species. The northern and southern areas of the bay show minimum values of specific richness, diversity and equitability. Six and a half percent of the species (*M. furnieri*, and *M. ancylodon*) are **permanent** ones (*i.e.* presence >75%); 48,4% are **frequent** species (between 10% and 50% of species, *B. aurea*, *A. marini*, *M. platanus*, *Odontesthes* sp., *P. brasiliensis*, *P. signata*, *P. cromis*, *P. valenciennesi*, *flatfishes*, *M. americanus*, *S. brasiliensis*, *C. striatus*, *M. goodei*, *C. carpio* and *L. grossidens*), and el 43.1% are **occasional species** (presence <10%, *P. nudigula*, *P. saltatrix*, *U. canosai*, *Astyanax* spp, *Corydoras* sp., *Acanthistius brasilianus*, *Engraulis anchoita*, *G. barbuis*, *P. pororosissimus*, *S. plagiusa*, *Ramnogaster arcuata*, *P. albicans* and *P. maculatus*). Marine species (*P. signata* and flounders of the genera *Paralichthys* and *Xystreurys*) and some freshwater species (*P.valenciennesi* and *C. carpio*) co-exist within a salinity range from 12 to 14 SPU. When salinity values are higher or lower, there are present only strictly marine or freshwater species respectively (Lasta 1995).

The seasonal variation in the specific composition within Samborombón Bay appears related to the spatial extension of the fish assemblages of the estuary which depends of the bi-modal discharge pattern of the Río de la Plata (Jaureguizar et al 2003, 2004, 2006). There also changes due to seasonal migrations.

Brevoortia aurea and *P. cromis* spawn in the bay (Lasta 1995, Macchi et al. 2002), which is the main reproductive area of resident species as *B. aurea* and *M. ancyloдон* and migratory ones as *M. furnieri* and *Mugil* sp. *A. marinii* and *Odontesthes* sp. (8.8% of the species) developed all their life cycles within the Bay. Other, as *M. furnieri*, *M. ancyloдон* and *M. platanus*, have in the Bay zones of larvae and breeding concentration. There is a large percentage of species (*P. brasiliensis*, *G. barbuis*, *P. signata*, flatfish, *M. americanus*, *P. cromis*, *U. brasiliensis*, *C. orbignyanus*, *P. valenciennesi*, *S. plagiusa*, *P. albicans* and *P. maculatus*), that appear as juvenile and adult but do not reproduce in the Bay (Lasta 1995).

2. Fish assemblages from Río de la Plata and its Maritime Front

2.1. Fish assemblages from the river to the slope

The Río de la Plata and its Marine Front ichthyofauna has a strong spatial arrangement along the environment gradient from the fresh and shallow water (3.5 m) to marine deeper water (323.5 m). The species composition, defines four main areas: riverine, estuarine, shelf and slope (Fig. 1). These areas, consistently distinguishable by their environmental conditions (Table 2) and consistent

boundaries, evidence a gradual change from riverine to marine communities rather than a distinct single transition (Fig. 2) (García et al. 2010).

The riverine area covers the inner part of the Río de la Plata and is characterized by fresh water, shallow depth and the highest water temperature (Fig. 1, Table 2). The fish composition has the highest similarity, and is mainly dominated by freshwater species (*P. vetula*, *P. albicans*, *L. obtusidens*, *P. granulatus*, *C. carpio*, and *L. pati*), and anadromous species (*G. barbus* and *L. grossidens*) (Fig. 2). The anadromous species, *G. barbus* during spring and early summer and *L. grossidens* during the autumn and early winter, penetrates into the Río de la Plata and rivers from the Río de la Plata basin to spawn (Fuster de Plaza and Boschi 1961, Ringuélet et al. 1967, Ringuélet 1975, Jaureguizar et al. 2003, Menni 2004).

In the Estuarine area (Fig. 1), with intermediate salinities and temperatures (Table 2), the fish community has an intermediate internal similarity associated to predominance of estuarine resident species (*M. furnieri*, *A. marinii*, *B. aurea*, *P. brasiliensis*, and *M. ancylodon*), and to a lower degree to occasional freshwater species (*P. albicans*, *P. valenciensis*, and *L. pati*) and marine species, either straggler (*C. guatucupa*, *C. orbignyanus* and *P. patagonicus*) or migrant (*P. punctatus* and *S. bonapartii*) (Fig. 2). The estuarine resident species use Samborombón Bay as the main nursery area (Lasta 1995). The straggler species (*C. guatucupa* and *P. patagonicus*) occurred more frequently in shelf waters, and carried on reproductive activity in coastal zones with salinity over 28 psu (Macchi and Acha 1998).

The area adjacent to the Río de la Plata (salty, intermediate depth and cool) form the shelf area (Fig. 1, Table 2), where the ichthyofauna has an intermediate

internal similarity, and is mainly dominated by marine species, namely *T. lathami*, *M. schmitti*, *S. guggenheim*, *P. brasiliensis*, *P. nudigula*, *S. brasiliensis*, *M. argentinae*, *P. pagrus*, *M. furnieri* and *D. tschudii*.

The slope area covers the zone with the greatest depths, highest salinities, and the lowest temperatures (Fig. 1, Table 2) and its fish community showed the lowest internal similarity. In contrast with the shelf area, the slope assemblage is dominated by marine species associated with deep water (*S. acanthias*, *M. hubbsi*, *M. magellanicus*, *H. lahillei* and *B. albescens*) (Fig. 2).

This fish assemblage pattern is mainly influenced by the bottom salinity and temperature, and each area shows consistent boundaries which occur near the location of frontal zone (García et al. 2010) (Fig. 2). The riverine–estuarine border occurs where the halocline intersects the bottom, and corresponds to the bottom salinity front defined by Guerrero et al. (1997a, b). This represents a frontier for intrusions of freshwater species into the estuary (*L. pati*, *P. valenciennis*, *L. obtusidens*, *P. lineatus* and species of Loricariidae) (Boschi 1988, Jaureguizar et al. 2003). The border zone between the estuarine and shelf areas coincides with the location of the maximum horizontal gradient of surface salinity (Guerrero et al. 1997a, b), indicating the boundary between the estuary and the continental coastal waters (Mianzan et al. 2001). The persistent salinity vertical structure (Guerrero et al. 1997a, b) in the estuarine area (Fig. 2), shows the presence of a shelf water intrusion along the bottom, which allows the incursion of marine species toward the head of the estuary (Jaureguizar et al. 2003). This zone is the limit for the presence in the estuary of marine species as *M. argentinus*, *P. pagrus*, *M. hubbsi* and *T. lathami*. (Boschi 1988, Jaureguizar et al. 2003). The shelf-break front is near the

border between shelf and slope fish assemblages. This front is a permanent feature that characterized the border of the shelf, and its inner boundary lies between 90 and 100 m isobaths (Acha et al. 2004). Here the sub Antarctic waters meet the cooler and more saline water of the Malvinas current, thus producing a thermohaline front (Lutz and Carreto 1991, Martos and Piccolo 1988).

The BIO-ENV process analysis identified surface and bottom salinity ($\rho = 0.848$, and $\rho = 0.819$ respectively) as the factors having the greatest influence on the fish assemblages distribution based on biomass. Bottom temperature ($\rho = 0.828$) was the next most influential factor on the fish assemblages distribution based on abundance. For both analyses the combination of temperature and bottom salinity present the best correlation coefficient ($\rho = 0.906$ and 0.917 respectively).

2.2. Fish assemblage composition in relation to feeding

Considering the species feeding, the fish assemblage composition reflects a transition from a fish community influenced by muddy deposits originated in the river, dominated by bottom or detritus feeders, to a fish community where the presence of marine pelagic or semi pelagic species becomes more important, where probably energy flow originating from bottom deposits is less important. In the second case, species feeding on small pelagic fish or crustaceans indicate that the energy derived from the phytoplankton and zooplankton production becomes more important. Most of the freshwater species are bottom or bottom related species, omnivorous and euriphagic (Ringuelet 1975, Menni 2004). *Pterodoras granulosus* (Panattieri and del Barco 1982, Darrigran and Colautti 1994, Ferriz et al. 2000) and *L. obtusidens* (Mastrarrigo 1950, Ringuelet et al. 1967) and have a

similar diet. Due to their euriphagy, all these species have changed their diet because of the invasion of the Río de la Plata basin by the south eastern Asia molluscs *Corbicula fluminea* and *Limnoperna fortunei*, (Ferriz et al. 2000, García and Protogino 2005, García and Moltalto 2006). *Genidens barbatus* is a benthophagous species. Juveniles of *L. grossidens* are planktophagous and adults are ichthyophagous (Ringuelet et al. 1967). Most of species in the estuarine area have benthic habits and feed on molluscs and crustaceans. *M. furnieri* adults mainly prey upon *Macra isabelleana* and secondarily upon shrimps, cephalopods and polychaetes (Sánchez et al. 1991). Only *B. aurea* is planktophagous, preying on diatoms, dinoflagellates, and copepods (Sánchez 1999), but detritus has also been observed in its diet (Giangiobbe and Sánchez, 1993). In the shelf and slope, the feeding habits show a transition from a benthic diet, basically composed by molluscs, crustaceans and small fish, to a more benthic-pelagic diet composed by small pelagic fish and deep water crustaceans and occasionally zooplankton. Among fishes of these communities, *S. acanthias* prefers to prey on pelagic communities. The main food items are ctenophores, teleost fishes (*M. hubbsi*, *S. brasiliensis*, *E. anchoita*, *N. bergi*, notothenids and mictophids and cephalopods, and occasionally, epibenthic macrocrustaceans (Menni, 1985; García de la Rosa and Sánchez, 1997). *Mustelus schmitti* feed on crustaceans, polychaetes and fishes (Olivier et al. 1968, Menni, 1985). The principal dietary component of *S. guggenheim* are bony fish, followed by crustaceans, shrimps, molluscs and polychaetes. The bony fish are mainly pelagic fishes such as *E. anchoita*, followed by demersal fishes such as *C. guatucupa* and *P. ramsayi*, *N. longipes* and *M. hubbsi* (Cousseau 1973, Vögler et al. 2003). *Helicolenus lahillei* feeds on benthic

and pelagic species, such as ctenophores, salps, crustaceans, squids and fishes (Cousseau and Perrotta 2000). Adult *M. hubbsi* mainly feed on pelagic and demersal-pelagic species, with the more common prey being zooplanktonic crustaceans amphipods and euphausiids, cephalopods, fishes, and epibenthic microcrustaceans (Angelescu et al. 1958, Angelescu and Cousseau 1969, Angelescu and Prenski 1987, Ruiz and Fondacaro 1997, Cordo 1981, Sánchez and García de la Rosa 1999). Juveniles mainly prey on zooplankton and secondarily on cephalopods and epibenthic macrocrustaceans. *Trachurus lathami* feed on copepods and chaetognaths (Cousseau 1967). *Percophis brasiliensis* feed on fishes and squids (San Román 1972). *Macruronus magellanicus* is an eurybatic species, able to adapt to different depths and hydrological conditions. It feeds upon pelagic crustaceans, amphipods, cephalopods and small fishes like sardines and notothenids (Angelescu and Gneri 1961, Bezzi 1984, Sánchez and Prenski 1996, Sánchez 1999).

Besides the important role played by salinity and temperature in habitat differentiation, the change in the food energy source from benthic production or detritus to phytoplankton and zooplankton evidence their importance in structuring demersal fish communities.

3. Assessing the genetic structure of fish populations

The genetic population structure of a fish species within a particular geographic area may take different forms. In fisheries biology the stock, rather than the local population, is frequently referred to as the basic unit for harvest and management. The stock concept should be redefined to be synonymous of a grouping that

relates to reproductive relationships and has a genetic meaning (such as local population or genetic patch). Biologically sustainable management based on knowledge of this structure may reduce the risk for depletion of genetic resources (Laikre et al. 2005).

Up to date, most empirical population genetic studies have been based on genetic markers assumed to primarily reflect selectively neutral DNA variation (most allozymes, microsatellites and mtDNA markers). Recent molecular technique developments provide increased opportunities for studying markers known to be located within functional genes of potential importance for fitness (Cano et al. 2008).

3.1. Recent studies in population genetic structure in different ecotypes of fish of the Río de la Plata and its Maritime Front

3.1.1. Pelagic fish

Despite the lack of genetic differentiation on a large geographic scale, many studies on pelagic fish detected small but statistically significant genetic differentiation among samples collected from very proximate localities within a short interval of time (Grande 1985).

Brevoortia aurea. This is a migratory pelagic marine fish from the Southwestern Atlantic and an abundant commercial fish in the Río de la Plata and its Maritime Front. The Brazilian menhaden represents an important species model to investigate patterns of genetic differentiation. Based on *cyt-b* phylogeographic approach, García et al. (2008) showed unexpected high genetic structuring in this pelagic marine fish. **The analyses suggested the existence of a long-term stability**

of the *B. aurea* schools, leading to a microgeographical genetic differentiation after the mixture of different stocks in the Río de la Plata and in the Atlantic Ocean from the associated estuarine environments. The data revealed that the recruitment of unrelated mtDNA haplotypes carried out by individuals within schools could be occurring in the same nursery areas, revealing the existence of many different maternal lineages. The AMOVA analysis revealed a high level of intrapopulation diversity consistent with an extensive and low regional structured population in *B. aurea*. The highest within-population estimates of gene diversity (Fig. 3, Table 3. a) found in the *cyt-b* data set of *B. aurea* could be also congruent with a scenario of a metapopulation, where floods generated by the opening of the sand bars in rivers and lagoons mix individuals across large distances.

Engraulis anchoita. The anchovy is widely distributed in tropical and sub-tropical waters of the southwestern Atlantic and spawn in open coastal areas in the inner continental shelf. Adults move during seasons between open coastal areas and bays, where they form large aggregations targeted by important fisheries (Araújo et al. 2008).

Present phylogeographic analysis based on cytochrome b sequences in *E. anchoita* from four different collecting regions (Fig. 3) revealed unexpected large values for haplotype diversity and nucleotide diversity (Table 3b). Phylogenetic reconstruction using neighbour joining tree based on HKY85 model of molecular substitution (Fig. 3) showed two major clusters of taxa. One major group included most and diverse haplotypes belonging to the inner continental shelf (at 35°S-53°W). The other one was integrated by haplotypes from the outer Río de la Plata, outer shelf and inner shelf (at 36°S-54°W). In the AMOVA analysis two grouping of

samples hypothesis (outer Río de la Plata and inner shelf at 36°S-54°W vs. Outer shelf and inner continental shelf at 35°S-53°W) have obtained the largest percentage of molecular variance. This grouping hypothesis showed significant values of the among groups component (Φ_{CT}) of the molecular variance. These results agree with the presence of at least two different long-term genetic stocks of *E. anchoita* in the Río de la Plata and its Maritime Front. *E. anchoita* moves around southern Uruguay and northeastern Argentina from offshore toward the end of the year and northward in March to June (Csirke 2005). Differences among growth rates of anchovy larvae from Brazilian Southeastern Bight suggest that this taxon may conform a distinct population from more southerly stocks (Castello and Castello 2003). Therefore the mix of different stocks in the border of the Río de la Plata estuary and Southwestern Atlantic Ocean may explain the high level of genetic diversity detected in *E. anchoita*.

3.1.2. Benthic fish

Squatina guggenheim. This is a benthic elasmobranchs that inhabits shelf and upper slope environments in temperate and tropical regions of the world (Compagno 1984). This species is one of the main coastal fishing resources having a wide geographic distribution from Espírito Santo (Brazil) to central Patagonia, Argentina (Vooren and da Silva 1991), in waters 10–80 m deep (Cousseau and Figueroa 2001). Like other benthic elasmobranchs, it tends to have low dispersal capability, which usually results in specimens from nearby areas having almost no mixing. As this restricted mixing may produce different life history parameters, it is important to study possible life history differences within angel shark species, even at small geographic scales (Colonello et al. 2007).

Phylogeographic approach based on cytochrome b sequences in *S. guggenheim* from three different collecting regions revealed very low values of haplotype and nucleotide diversity (Table 3c). The phylogenetic reconstruction by using neighbour joining method based on GTR model of molecular substitution (Fig. 4) shows three major cluster. One well supported group integrated by haplotypes from the outer Río de la Plata and outer shelf, a second one conformed by samples from Atlantic Ocean coast and outer Río de la Plata, and the last one integrated only by different Atlantic Ocean coast haplotypes. Remarkably the most frequent haplotype 1 was shared by samples from the three aforementioned regions. Different grouping hypotheses performed in the AMOVA analysis failed to detect significant values in the geographic partition of the molecular variance (García et al. 2009). Though a high population structuring was expected, all present data support the hypothesis that *S. guggenheim* constitutes a single panmictic unit or genetic stock in the Río de la Plata and its Maritime Front.

3.1.3 Demersal fish

Micropogonias furnieri. Populations of this species are distributed from the southern Caribbean to the Gulf of San Matías in Argentina. The white croaker is an important commercial fishing resource and at present it is considered to be fully exploited. A population genetic analysis based on mitochondrial D-loop region (CR) (Pereira et al. 2009) detected the presence of two stocks of *M. furnieri* in three different areas (Bahía Blanca, Río de la Plata and Atlantic Ocean). Río de la Plata locality have the lower values of haplotype and nucleotide diversity as well as demographic signatures of population declination whereas Atlantic Ocean

collecting sites showed the highest values of genetic diversity and signatures of a past recent population expansion.

3.2. Units for fishery management, conservation and potential effects of ignoring the structure.

Present population genetic data indicate a long-term stocks differentiation between Río de la Plata and the Atlantic Ocean environments since the Pleistocene both in *B. aurea* (García et al. 2008) and *M. furnieri* (Pereira et al. 2009). In *B. aurea* the analysis suggests that the metapopulation represents the management unit, as well as the need of the inclusion of the associated rivers and coastal lagoons which are responsible for the origin of the major genetic diversity, in a high priority conservation program within a Marine Protected Area System in the Southwestern Atlantic Ocean (García et al. 2008). Biologically improper management may result in overharvest of particular populations or population segments, resulting in extinction or reduction of genetic diversity within populations. In this sense the smallest population sizes and the lowest levels of genetic variability in *M. furnieri* from the Río de la Plata, make this stock vulnerable to over-exploitation (Pereira et al. 2009).

Though Csirke (2005) have suggest that *E. anchoita* could support a higher fishing pressure, present analyses claims for a stocks management as mechanism to retain the basal level of genetic diversity in this taxon.

Low level of population structure was evident in *S. guggenheim*. The lowest genetic diversity values detected in this taxon could be indicating the existence of a

long-term population declination or a past recent population bottleneck making it highly vulnerable to overexploitation (García et al. 2009).

BIBLIOGRAPHY

- Abella A, Arena G, Nion H, Ríos C (1979) Peces bentónicos del Río de la Plata y de la zona común de pesca Argentino-Uruguaya. In: Memorias del Seminario Ecología bentónica y sedimentación de la plataforma continental del Atlántico Sur. UNESCO, Montevideo, Uruguay, pp 291-324
- Acha M, Mianzan H (2003) El Estuario del Plata: donde el río se encuentra con el mar. Rev. Ciencia Hoy 13 (73):10-20
- Acha M, Mianzan HW, Guerrero RA, Favero MF, Bava J (2004) Marine fronts at the continental shelves of austral South America, Physical and ecological processes. J. Mar. Syst. 44: 83-105
- Almirón AE, Gómez SE, Toresani NI (1992) Peces de agua dulce de la provincia de Buenos Aires, Argentina. Situación ambiental de la Provincia de Buenos Aires. A. Recursos y rasgos naturales en la evaluación ambiental, CIC, La Plata Argentina 2(12):1-29
- Almirón AE, García ML, Menni RC, Protogino LC, Solari L (2000) Fish ecology of a seasonal lowland stream in temperate South America. Mar Freshwater Res 51 (3):265-274
- Angelescu V, Cousseau MB (1969) Alimentación de la merluza en la región del talud continental argentino, época invernal (Merluccidae, *Merluccius*, *Merluccius hubbsi*). Bol Inst Biol Mar, Mar del Plata 19:1-93
- Angelescu V, Gneri FS, Nani A (1958) La merluza del Mar Argentino. Secret Mar Serv Hidrog Naval H 1004:1-224

- Angelescu V, Gneri FS (1961) Contribución al conocimiento bioecológico de la merluza de cola (*Macruronus magellanicus* Lönngberg). Actas I Cong Sudam Zool 1:3-18
- Angelescu V, Prenski LB (1987) Ecología trófica de la merluza común del Mar Argentino, (Merluccidae, *Merluccius hubbsi*). Parte II. Dinámica de la alimentación analizada sobre la base de las condiciones ambientales, la estructura y las evaluaciones de los efectivos en su área de distribución. Contr INIDEP Mar del Plata 561:1-205
- Araújo FG, Silva MA, Santos JNS, Vasconcellos RM (2008) Habitat selection by anchovies (Clupeiformes: Engraulidae) in a tropical bay at Southeastern Brazil. Neotropical Ichthyology 6:583-590
- Azpelicueta MM, Almirón AE (1999) A sturgeon (Acipenseridae) in temperate waters of the South Hemisphere, Río de la Plata, Argentina. Biogeographica 75(3):129-130
- Bezzi SI (1984) Aspectos biológicos pesqueros de la merluza de cola del Atlántico sudoccidental. Rev Inv Des Pesq 4: 63-80
- Boschi EE (1988) El ecosistema del Río de la Plata (Argentina y Uruguay). Anal Inst Cien Mar y Limnol UNAM 15:159-182
- Cano JM, Shikano T, Kuparinen A, Merilä J (2008) Genetic differentiation, effective population size and gene flow in marine fishes: implications for stock management. JIFS 5:1-10
- Castello L, Castello JP (2003) Anchovy stocks (*Engraulis anchoita*) and larval growth in the SW Atlantic. Fish Res 59:409-421

- Colonello JH, Lucifora LO, Massa A (2007) Reproduction of the angular angel shark (*Squatina guggenheim*): geographic differences, reproductive cycle, and sexual dimorphism. ICES J Mar Sci 64:131–140
- Compagno LJV (1984) Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Part 1 FAO species catalogue 4 FAO Fisheries Synopsis 125:1–249
- Cordo HD (1981) Resultados sobre la alimentación de la merluza del Mar Epicontinental Argentino (*Merluccius hubbsi*). Análisis biológico y estadístico de los datos obtenidos de la campañas de B/I “Shinkai Maru” y “Walther Herwig” (1978-1979). In: Angelescu V (ed) Campañas de investigación pesquera realizadas en el Mar Argentino por los B/I “Shinkai Maru” y “Walther Herwig”, años 1978 y 1979. Resultados de la parte argentina. Contr INIDEP, Mar del Plata 383 pp 299-312
- Cousseau MB (1967) Contribución al conocimiento de la biología del surel (*Trachurus picturatus australis*) del área de Mar del Plata (*Pisces*, Fam. *Carangidae*). Bol Inst Biol Mar 15:1-38
- Cousseau MB (1973) Taxonomía y biología del pez ángel, *Squatina argentina* Marini (*Pisces*, Squatinidae). Physis 32:175-195
- Cousseau MB (1985) Los peces del Río de la Plata y su Frente Marítimo. In Yáñez-Arancibia A (ed) Estuaries and Coastal Lagoons: Towards an Ecosystem Integration, DR (R) UNAM Press, México, pp 515-534
- Cousseau MB, Figueroa DE (2001) Las especies del género *Squatina* en aguas de Argentina (*Pisces*: Elasmobranchii: Squatinidae). Neotrópica 47:85–86

- Cousseau MB, Perrota RG (2000) Peces marinos de Argentina. Biología, distribución, pesca. INIDEP, Mar del Plata
- Csirke J (2005) Review of the state of world marine fishery resources. FAO fisheries. Technical Paper 457. Publishing Management Service Rome Italy.
- Darrigran GA, Colautti D (1994) Potencial control biológico del molusco invasor *Corbicula fluminea* (Müller, 1774) en el Río de la Plata. Comunic Soc Malacol Uruguay 7:368-373
- De Buen F (1950) El Mar de Solís y su fauna de peces (2da. Parte). Serv Oceanog Pesca del Uruguay, 2:45-144
- Ferriz RA, Villar CA, Colautti D, Bonetto C (2000) Alimentación de *Pterodoras granulosus* (Valenciennes) (Pisces, Doradidae) en la baja cuenca del Plata. Rev Mus Arg Cienc Nat ns 2:151-156
- Fuster de Plaza ML, Boschi EE (1961) Áreas de migración y ecología de la anchoa *Lycengraulis olidus* (Günther) en las aguas argentinas (Pises, fam. Engraulidae). Bol Inst Biol Mar 1:1-58
- García G, Vergara J, Gutiérrez V (2008) Phylogeography of the Southern Atlantic menhaden *Brevoortia aurea* inferred from mitochondrial cytochrome b gene. Mar Biol 155:325–336
- García G, Pereyra S, Oviedo S, Miller P, Domingo A (2009) Estructura genética del angelito (*Squatina* spp.) en el Río de la Plata y su Frente Marítimo. In Proceedings of VII Simp Rec Genét Amér Lat y el Caribe 2 INIA Carrillanca, Pucón Chile, pp 173-174

- García M L, Moltalto L (2006) Impacto en el ambiente natural. In: Darrigran G, Damborenea C (eds.) Bio-invasión del mejillón dorado, *Limnoperna fortunei*, en el continente americano. Edulp, La Plata, pp:111-127
- García ML, Protogino LC (2005) Invasive freshwater molluscs are consumed by native fishes in South America. *J Appl Ichthyol* 21:34-38
- García M, Jaureguizar A, Protogino L (2003) Fish assemblages along a riverine-marine environment gradient. <http://www.freplata.org/documentos/otros.asp>.
- García M, Jaureguizar A, Protogino L (2010) From fresh water to the slope: fish community ecology in the Río de la Plata and the sea beyond. *Lat Am J Aquat Res* 38(1):81-94
- García de la Rosa SB, Sánchez F (1997) Alimentación de *Squalus acanthias* y predación sobre *Merluccius hubbsi* en el Mar Argentino entre 34°50'-47°S. *Rev Inv Des Pesq* 11:119-133
- García Romero N, Azpelicueta MM, Almirón AE, Casciotta RJ (1998) *Hypophthalmichthys molitrix* (Cypriniformes: Cyprinidae). Other exotic cyprinid in the Río de la Plata. *Biogeographica* 74 (4):189-191
- Giangiobbe A, Sánchez F (1993) Alimentación de la saraca (*Brevoortia aurea*). *Frente Marítimo* 14:71-80
- Grande L (1985) Recent and fossil clupeomorph fishes with materials for revision of the subgroups of clupeoids. *Bull Am Mus Nat Hist*:235-372
- Guerrero RA, EM Acha, MB Framiñan, CA Lasta (1997a) Physical oceanography of the Río de la Plata estuary, Argentina. *Cont Shelf Res* 17:727-742
- Guerrero RA, Lasta CA, Acha EM, Mianzan H, Framiñan M (1997b) Atlas hidrográfico del Río de la Plata. CARP-INIDEP, Mar del Plata, Argentina

- Huelsenbeck JP, Crandall KA (1997) Phylogeny estimation and hypothesis testing using maximum likelihood. *Annu Rev Ecol Syst* 28:437–66
- Jaureguizar AJ, Waessle JA, Guerrero RA (2007) Estuarine dynamics controlling the Atlantic searobins (*Prionotus* spp) distribution on Southwestern Atlantic Coastal System (34°-41° S). *Estuar Coast Shelf Sci* 73: 30-42
- Jaureguizar AJ, Menni R, Guerrero R, Lasta C (2004) Environmental factors structuring fish communities of the Río de la Plata estuary. *Fish Res* 66:195-211
- Jaureguizar AJ, Menni R, Lasta C, Guerrero R (2006) Fish assemblages of the northern Argentine coastal system: spatial patterns and their temporal variations. *Fisheries Oceanogr* 15:326-344
- Jaureguizar AJ, Menni RC, Bremec C, Mianzan H, Lasta C (2003) Fishes assemblage and environmental patterns in the Río de la Plata estuary. *Estuar. Coast Shelf Sci* 56:921-933
- Laikre L, Palm S, Ryman N (2005) Genetic Population Structure of Fishes: Implications for Coastal Zone Management. *Ambio* 34:111-119
- Lasta CA (1995) La Bahía Samborombón: zona de desove y cría de peces. Tesis Doctoral, Facultad de Ciencias Naturales, Universidad Nacional de La Plata, Argentina
- Liotta J, Giacosa B, Wagner M (1995/1996) Lista comentada de la ictiofauna del Delta del Paraná. *Rev Ictiol* 4(1-2):23-32
- López HL, Miquelarena AM, Menni RC (2003) Lista comentada de los peces continentales de la Argentina. *Ser Téc Didác ProBiota* 5, pp 86

- López HL, Menni RC, Ferris, Ponte Gómez J, Cuello MV (2006) Bibliografía de los peces continentales de la Argentina. <http://www.fcgyn.unam.edu.ar/icades>.
<http://aquacomm.fcla.edu/1668>
- López, HL, Menni RC, Donato M, Miquelarena AM (2008) Biogeographical review of Argentina (Andean and Neotropical regions): an análisis using freshwater fishes. *J Biogeogr* 1-16.
- López HL, Morgan MC, Montenegro MJ (2002) Ichthyological ecoregions of Argentina. Documents Series, Probiota, La Plata
- López HL, Protogino LC, Aquino AE (1996) Ictiología continental de la Argentina: Santiago del Estero, Catamarca, Córdoba, San Luis, La Pampa y Buenos Aires. *Aquatec* 3:1-14
- Lutz VA, Carreto JI (1991) A new spectrofluorometric method for the determination of chlorophylls and degradation products and its application in two frontal areas of the Argentine Sea. *Cont Shelf Res* 11:433-451
- Macchi GJ, Acha EM (1998) Aspectos reproductivos de las principales especies de peces en la Zona Común de Pesca Argentino-Uruguayo y en el Rincón. Noviembre, 1994. In: Lasta CA (ed) Resultados de una campaña de evaluación de recursos demersales costeros de la Provincia de Buenos Aires y del litoral Uruguayo. INIDEP, Mar del Plata, pp 67-89
- Mac Donagh EJ (1945) Pesca de una “carpa de espejuelos” en el Río de la Plata. *Not Mus La Plata X Zool* 69:315-324
- Marelli CA (1924) Elenco sistemática de la fauna de la provincia de Buenos Aires (Procordados y Vertebrados). *Mem Min Obras Públ* 1922-1923:536-682

- Martos P, Piccolo MC (1988) Hydrography of the Argentine continental shelf between 38° and 42°S. *Cont Shelf Res* 8:1043-1056
- Mastrarrigo V (1950) La Boga. Contribución a su conocimiento biológico. *Almanaque Minist Agric y Ganad* 25: 417-426.
- Menni RC (1985) Distribución y biología de *Squalus acanthias*, *Mustelus schmitti* y *Galeorhinus vitaminicus* en Agosto-Setiembre de 1978 en el Mar Argentino (Chondrichthyes). *Rev Mus La Plata NS* 13 138:151-202
- Menni RC (2004) Peces y ambientes en la Argentina continental. *Monog Mus Arg Cienc Nat* 5:1-316
- Menni RC, Ringuelet RA, Arámburu RH (1984) Peces marinos de la Argentina y Uruguay. Hemisferio Sur, Buenos Aires
- Mianzan H, Lasta CA, Acha EM, Guerrero RA, Macchi G, Bremec C (2001) The Río de la Plata estuary, Argentina-Uruguay. In: Seeliger U, de Larceda LD, Kjerve B (eds). *Ecological studies: coastal marine ecosystems of Latin America*. Springer, Berlin, pp. 185-204
- Nei M (1987) *Molecular Evolutionary Genetics*. Columbia University Press, New York
- Nion H (1998) Peces del Río de la Plata y algunos aspectos de su ecología. In: Wells PG, Daborn GR (eds), *El Río de la Plata. Una revisión ambiental*. University of Dalhousie, Canadá, pp. 169-190
- Olivier S, Bastida R, Torti MR (1968) Ecosistemas de aguas litorales. *Serv Hidrogr Naval H* 1025:1-45
- Panattieri AE, del Barco D (1982) Peces de la provincia de Santa Fe. Peces omnívoros, preferentemente de fondo, de verano. *Armado gallego*

- (*Pterodoras granulosus*). Cienc Tecnolg Agrop Minist Agric Ganad Prov Santa Fe 25:21-23
- Pereira AN, Márquez A, Marin M, Marin Y (2009) Genetic evidence of two stocks of the whitemouth croaker *Micropogonias furnieri* in the Río de la Plata and oceanic front in Uruguay. J Fish Biol 75:321–331
- Popovici Z, Riggi AE (1948) Los estudios de hidrobiología en la Argentina. Sus relaciones con el plan del superior gobierno de la nación y sus proyecciones futuras. Mus Arg C. Nat “B. Rivadavia” Misc 1:i-ix:1-171
- Posada D, Crandall A (1998) Modeltest: testing the model of DNA substitution. Bioinformatics 14:817-818
- Pozzi A J (1945) Los peces de agua dulce de la República Argentina. GAEA, An Soc Arg Est Geogr Argentina T VII:239-292
- Ringuelet RA (1962) Ecología acuática continental. EUDEBA Buenos Aires
- Ringuelet RA (1975) Zoogeografía y ecología de los peces de aguas continentales de la Argentina y consideraciones sobre las áreas ictiológicas de América del Sur. Ecosur 2 (3):1-122
- Ringuelet RA, Arámburu RH (1957) Enumeración sistemática de los vertebrados de la provincia de Buenos Aires. Min Asunt Agr Prov Buenos Aires 19:7-25
- Ringuelet RA, Arámburu RH, de Arámburu AA (1967) Los peces argentinos de agua dulce. Com. Invest Cient Prov Buenos Aires, La Plata
- Ruiz AE, Fondacaro RR (1997) Diet of hake (*Merluccius hubbsi* Marini) in a spawning and nursery area within Patagonian shelf waters. Fish Res 30:157-160

- Sánchez F (1999) Ecología trófica de la merluza de cola (*Macruronus magellanicus*) del Atlántico sudoccidental. In: Proyecto INIDEP-JICA. Avances en métodos y tecnología aplicados a la investigación pesquera. Seminario final del Proyecto INIDEP-JICA sobre evaluación y monitoreo de recursos pesqueros 1994-1999. Inst. Nac. Inv. Des. Pesq. 135-138
- Sánchez F, García de la Rosa SB (1999) Alimentación de *Merluccius hubbsi* e impacto del canibalismo en la región comprendida entre 34°50'-47°S del Atlántico sudoccidental. Rev Inv Des Pesq 12:77-93
- Sánchez F, Prenski LB (1996) Ecología trófica de peces demersales en el golfo San Jorge. Rev Inv Des Pesq 10:57-71
- Sánchez F, Mari N, Lasta C, Giangiobbe A (1991) Alimentación de la corvina rubia (*Micropogonias furnieri*) en la Bahía Samborombón. Frente Marítimo 8:43-50
- San Román N, (1972) Alimentación del "pez palo", *Percophis brasiliensis* Quoy and Gaimard, 1824. Physis 31:605-612
- Vögler R, Milessi AC, R.A.Quiñones RA (2003) Trophic ecology of *Squatina guggenheim* on the continental shelf off Uruguay and northern Argentina. J Fish Biol 62:1254-1267
- Vooren CM, da Silva KG (1991) On the taxonomy of the angel sharks from southern Brazil, with the description of *Squatina occulta* sp. n. R Br Biol 51:589-602

Table 1. Fish species from the Río de la Plata and its Maritime Front. FS: freshwater species, BS: brackish species, MS: marine species.

	FS	BS	MS		FS	BS	MS
Clase Chondrichthyes				<i>Myliobatis goodei</i>			X
Familia Odontaspidae				Familia Callorhynchidae			
<i>Carcharias taurus</i>			X	<i>Callorhynchus callorhynchus</i>			X
Familia Lamnidae				Clase Actinopterygii			
<i>Alopias vulpinus</i>			X	Familia Acipenseridae			
Familia Scyliorhinidae				<i>Acipenser cf. baerii</i>	X	X	
<i>Schoederichthys bivius</i>			X	Familia Congridae			
Familia Carcharhinidae				<i>Conger orbignyanus</i>		X	
<i>Carcharinus plumbeus</i>			X	<i>Pseudoxenomystax albescens</i>			X
Familia Triakidae				Familia Pristigasteridae			X
<i>Galeorhinus galeus</i>			X	<i>Pellona flavipinnis</i>	X		
<i>Mustelus canis</i>			X	Familia Engarulidae			
<i>Mustelus fasciatus</i>			X	<i>Anchoa marinii</i>		X	
<i>Mustelus schmitti</i>			X	<i>Engraulis anchoita</i>			X
Familia Hexanchidae				<i>Lycengarulis grossidens</i>		X	X
<i>Hexanchus griseus</i>			X	Familia Clupeidae			
<i>Notorhynchus cepedianus</i>			X	<i>Brevoortia aurea</i>		X	
Familia Sphyrnidae				<i>Platanichthys platana</i>	X	X	
<i>Sphyrna zygaena</i>				<i>Ramnogaster arcuata,</i>		X	
Familia Squalidae			X	<i>Ramnogaster melanostoma</i>	X		
<i>Squalus acanthias</i>				Familia Cyprinidae			
<i>Squalus megalops</i>			X	<i>Cyprinus carpio</i>	X	X	
<i>Squalus mitsukuri</i>			X	<i>Hypophthalmichthys molitrix</i>	X		
Familia Squatinidae			X	Familia Parodontidae			
<i>Squatina argentina</i>				<i>Apareiodon affinis</i>	X		
<i>Squatina guggenheim</i>			X	<i>Parodon nasus</i>	X		
Familia Torpedinidae			X	Familia Curimatidae			
<i>Discopyge tschudii</i>				<i>Cyphocharax platanus</i>	X		
Familia Rhinobatidae			X	<i>Cyphocharax saladensis</i>	X		
<i>Rhinobatos horkelii</i>				<i>Cyphocharax spilotos</i>	X		
<i>Zapteryx brevirostris</i>			X	<i>Cyphocharax voga</i>	X		
Familia Rajidae			X	<i>Potamorhina squamoralevis</i>	X		
<i>Atlantoraja castelnaui</i>				<i>Steindachnerina biornata</i>	X		
<i>Atlantoraja cyclophora</i>			X	<i>Steindachnerina brevipinna</i>	X		
<i>Atlantoraja platana</i>			X	Familia Prochilodontidae			
<i>Rioraja agasizi</i>			X	<i>Prochilodus lineatus</i>	X		
<i>Psammobatis bergi</i>			X	Familia Anostomidae			
<i>Psammobatis rudis</i>			X	<i>Abramites hypselonotus</i>	X		
<i>Sympterygia acuta</i>				<i>Leporinus acutidens</i>	X		
<i>Sympterygia bonapartii</i>			X	<i>Leporinus obtusidens</i>	X		
<i>Zearaja flavirostris</i>			X	<i>Leporinus striatus</i>	X		
Familia Dasyatidae			X	<i>Pseudanos trimaculatus</i>	X		
<i>Dasyatis pastinaca</i>				<i>Schizodon platae</i>	X		
Familia Potamotrygonidae			X	<i>Pseudanos trimaculatus</i>	X		
<i>Potamotrygon brachyura</i>				<i>Schizodon platae</i>	X		
<i>Potamotrygon hystrix</i>		X		Familia Crenuchidae			
<i>Potamotrygon motoro</i>		X		<i>Characidium rachovii</i>	X		
Familia Gymnuridae		X		Familia Gasteropelecidae			
<i>Gymnura altavela</i>				<i>Thoracocharax stellatus</i>	X		
Familia Myliobatidae			X	Familia Characidae			
<i>Myliobatis freminvillii</i>			X	<i>Aphyocharax anisitsi</i>	X		

<i>Aphyocharax dentatus</i>	X	<i>Corydoras aeneus</i>	X
<i>Brycon orbygnianus</i>	X	<i>Corydoras paleatus</i>	X
<i>Charax stenopterus</i>	X	<i>Corydoras undulatus</i>	X
<i>Cynopotamus argenteus</i>	X	<i>Hoplosternum littorale</i>	X
<i>Galeocharax humeralis</i>	X	Familia Loricariidae	
<i>Roeboides affinis</i>	X	<i>Hisonotus maculipinnis</i>	X
<i>Roeboides microlepis</i>	X	<i>Macrotocinclus flexilis</i>	X
<i>Cheirodon interruptus</i>	X	<i>Otocinclus vittatus</i>	X
<i>Heterocheirodon yatai</i>	X	<i>Brochiloricaria chauliodon</i>	X
<i>Serrapinnus calliurus</i>	X	<i>Farlowella hahni</i>	X
<i>Diapoma terofali</i>	X	<i>Loricaria apeltogaster</i>	X
<i>Pseudocorynopoma doriai</i>	X	<i>Loricariichthys anus</i>	X
<i>Metynnix otuquensis</i>	X	<i>Loricariichthys platymetopon</i>	X
<i>Mylossoma duriventre</i>	X	<i>Paraloricaria commersonoides</i>	X
<i>Piaractus mesopotamicus</i>	X	<i>Paraloricaria vetula</i>	X
<i>Pygocentrus nattereri</i>	X	<i>Ricola macrops</i>	X
<i>Serrasalmus marginatus</i>	X	<i>Rineloricaria felipponei</i>	X
<i>Serrasalmus maculatus</i>	X	<i>Rineloricaria lima</i>	X
<i>Poptella paraguayensis</i>	X	<i>Rineloricaria pareiacantha</i>	X
<i>Tetragonopterus argenteus</i>	X	<i>Rineloricaria thrissoceps</i>	X
<i>Triportheus nematurus</i>	X	<i>Sturisoma robustum</i>	X
Incertae sedis		<i>Hypostomus commersoni</i>	X
<i>Astyanax abramis</i>	X	<i>Hypostomus laplatae</i>	X
<i>Astyanax asuncionensis</i>	X	<i>Hypostomus microstomus</i>	X
<i>Astyanax eigenmanniorum</i>	X	<i>Hypostomus punctatus</i>	X
<i>Astyanax erythropterus</i>	X	<i>Rhinelepis strigosa</i>	X
<i>Astyanax fasciatus</i>	X	<i>Ancistrus cirrhosus</i>	X
<i>Bryconamericus iheringii</i>	X	Familia Pseudopimelodidae	
<i>Bryconamericus stramineus</i>	X	<i>Mycroglanis cottoides</i>	X
<i>Ctenobrycon alleni</i>	X	Familia Heptapteridae	
<i>Hyphessobrycon anisitsi</i>	X	<i>Heptapterus mustelinus</i>	X
<i>Hyphessobrycon luetkenii</i>	X	<i>Pimelodella gracilis</i>	X
<i>Hyphessobrycon meridionalis</i>	X	<i>Pimelodella laticeps</i>	X
<i>Markiana nigripinnis</i>	X		
<i>Oligosarcus hepsetus</i>	X	<i>Rhamdia quelen</i>	X
<i>Oligosarcus jenynsii</i>	X	Familia Pimelodidae	
<i>Oligosarcus oligolepis</i>	X	<i>Bergiaria platana</i>	X
<i>Salminus brasiliensis</i>	X	<i>Hemisorubim platyrhynchus</i>	X
Familia Acestrorhynchidae		<i>Hypophthalmus edentatus</i>	X
<i>Acestrorhynchus pantaneiro</i>	X	<i>Iheringichthys labrosus</i>	X
Familia Cynodontidae		<i>Luciopimelodus argentinus</i>	X
<i>Raphiodon vulpinus</i>	X	<i>Luciopimelodus pati</i>	X
Familia Erythrinidae		<i>Megalonema platanun</i>	X
<i>Hoplias malabaricus</i>	X	<i>Parapimelodus valenciennis</i>	X
Familia Lebiasinidae		<i>Pimelodus albicans</i>	X
<i>Pyrrhulina australis</i>	X	<i>Pimelodus argenteus</i>	X
Familia Aspredinidae		<i>Pimelodus brevis</i>	X
<i>Bunocephalus doriae</i>	X	<i>Pimelodus maculatus</i>	X
<i>Bunocephalus iheringii</i>	X	<i>Paulicea luetkeni</i>	X
Familia Trichomycteridae		<i>Pseudoplatystoma coruscans</i>	X
<i>Scleronema angustirostre</i>	X	<i>Pseudoplatystoma reticulatum</i>	X
<i>Parastegophilus maculatus</i>	X	<i>Sorubim lima</i>	X
Familia Callichthyidae		<i>Zungaro jahu</i>	X
<i>Callichthys callichthys</i>	X	Familia Doradidae	

<i>Oxydoras kneri</i>				<i>Poecilia vivipara</i>	X		
<i>Pterodoras granulosus</i>	X			Familia Zeidae			
<i>Rhinodoras dorbignyi</i>	X			<i>Zenopsis conchifer</i>			X
Familia Auchenipteridae				Familia Syngnathidae			
<i>Ageneiosus inermis</i>	X			<i>Hippocampus punctulatus</i>			X
<i>Ageneiosus militaris</i>	X			<i>Syngnathus folletti</i>		X	X
<i>Auchenipterus nigripinnis</i>	X			Familia Synbranchidae			
<i>Auchenipterus osteomystax</i>	X			<i>Synbranchus marmoratus</i>	X		
<i>Trachelyopterus albicrux</i>	X			Familia Scorpaenidae			
<i>Trachelypterus striatulus</i>	X			<i>Helicolenus lahillei</i>			X
Familia Ariidae				Familia Congiopodidae			
<i>Genidens barbatus</i>	X			<i>Congiopus peruvianus</i>			X
Familia Gymnotidae				Familia Triglidae			
<i>Gymnotus inaequilabiatus</i>	X			<i>Prionotus nudigula</i>		X	X
Familia Sternopygidae				<i>Prionotus punctatus</i>			X
<i>Eigenmannia trilineata</i>	X			Familia Polyprionidae			
<i>Eigenmannia virescens</i>	X			<i>Polyprion americanus</i>			X
Familia Rhamphichthyidae				Familia Serranidae			
<i>Rhamphichthys rostratus</i>	X			<i>Acanthistius brasiliensis</i>			X
Familia Hypopomidae				<i>Dules auriga</i>			X
<i>Hypopomus artedi</i>	X			<i>Epinephelus guaza</i>			X
Familia Apterodontidae				Familia Pomatomidae			
<i>Apterodontus albifrons</i>	X			<i>Pomatomus saltatrix</i>		X	X
Familia Macrouridae				Familia Carangidae			
<i>Coelorhynchus marinii</i>			X	<i>Parona signata</i>		X	X
Familia Merlucciidae				<i>Selene vomer</i>			X
<i>Macruronus magellanicus</i>			X	<i>Trachurus picturatus</i>			X
<i>Merluccius hubbsi</i>			X	<i>Trachinotus marginatus</i>			X
Familia Phycidae				<i>Trachurus lathami</i>		X	X
<i>Urophycis brasiliensis</i>		X	X	<i>Vomer setapinnis</i>			X
Familia Ophidiidae				Familia Gerreidae			
<i>Genypterus blacodes</i>			X	<i>Eucinostomus gula</i>			X
<i>Raneya fluminensis</i>			X	<i>Eucinostomus melanopterus</i>		X	
Familia Batrachoididae				Familia Sparidae			
<i>Porichthys porosissimus</i>			X	<i>Diplodus argenteus</i>			X
<i>Thalassophryne montevidensis</i>			X	<i>Pagrus pagrus</i>			X
<i>Triathalassotheia argentina</i>			X	Familia Haemulidae			
Familia Mugilidae				<i>Boridia grossidens</i>			X
<i>Mugil platanus</i>	X	X	X	Familia Sciaenidae			
Familia Atherinopsidae				<i>Cynoscion guatucupa</i>			X
<i>Odontesthes argentinensis</i>		X	X	<i>Cynoscion striatus</i>			X
<i>Odontesthes bonariensis</i>	X	X		<i>Macrodon ancylodon</i>		X	X
<i>Odontesthes perugiae</i>	X			<i>Menticirrhus americanus</i>		X	X
<i>Odontesthes humensis</i>	X			<i>Micropogonias furnieri</i>			X
<i>Odontesthes retropinnis</i>	X			<i>Pachyurus bonariensis</i>		X	
Familia Rivulidae				<i>Paralonchurus brasiliensis</i>		X	X
<i>Austrolebias bellotti</i>	X			<i>Plagioscion ternetzi</i>		X	
<i>Austrolebias elongates</i>	X			<i>Pogonias cromis</i>			X
<i>Austrolebias nigripinnis</i>	X			<i>Umbrina canosai</i>			X
Familia Anablepidae				Familia Mullidae			
<i>Jenynsia multidentata</i>	X	X		<i>Mullus argentinae</i>			X
Familia Poeciliidae				Familia Cheilodactylidae			
<i>Cnesterodon decemmaculatus</i>	X	X		<i>Cheilodactylus bergi</i>			X
<i>Phalloceros caudimaculatus</i>	X			Familia Cichlidae			
<i>Phalloptychus januarius</i>	X			<i>Australoheros facetus</i>		X	

<i>Crenicichla lacustris</i>	X	<i>Trichiurus lepturus</i>		X
<i>Crenicichla lepidota</i>	X	Familia Scombridae		
<i>Gymnogeophagus australis</i>	X	<i>Scomber japonicus</i>		X
<i>Gymnogeophagus gymnogenys</i>	X	Familia Stromateidae		
<i>Gymnogeophagus meridionalis</i>	X	<i>Peprilus paru</i>		X
<i>Gymnogeophagus rhabdotus</i>	X	<i>Stromateus brasiliensis</i>	X	X
Familia Nototheniidae		Familia Paralichthyidae		
<i>Patagonotothen ramsayi</i>	X	<i>Etropus longimanus</i>		X
Familia Pinguipedidae		<i>Paralichthys isosceles</i>		X
<i>Pinguipes brasilanus</i>	X	<i>Paralichthys orbignyanus</i>	X	X
<i>Pseudopercis semifasciata</i>	X	<i>Paralichthys patagonicus</i>	X	X
Familia Percophidae		<i>Xystreuris rasile</i>		X
<i>Percophis brasiliensis</i>	X	Familia Pleuronectidae		
Familia Uranoscopidae		<i>Oncopterus darwini</i>	X	X
<i>Astroscopus sexspinosus</i>	X	Familia Achiriidae		
Familia Blenniidae		<i>Catathyridium jenynsii</i> ,	X	
<i>Hypleurochilus fissicornis</i>	X	Familia Cynoglossidae		
Familia Gobiidae		<i>Symphurus jenynsii</i>		
<i>Gobiosoma parri</i>	X	<i>Symphurus plagiusa</i>		
Familia Sphyraenidae		Familia Balistidae		
<i>Sphyraena picudilla</i>	X	<i>Balistes capriscus</i>		X
Familia Gempylidae		Familia Tetraodontidae		
<i>Thyrsitops lepidopodea</i>	X	<i>Lagocephalus laevigatus</i>		X
Familia Trichiuridae				

Table 2. Oceanographic data (mean, standard deviation) of fish assemblages areas defined by multivariate analyses using biomass (t nm⁻²) or abundance (thousands ind nm⁻²). Z, total depth (m); ST, surface temperature (°C); BT, bottom temperature (°C); SS, surface salinity (SPU); BS, bottom salinity (SPU).

		Riverine	Estuarine	Shelf	Slope
Biomass	Z	7.88 ± 1.53	6.29 ± 2.69	28 ± 12.75	151.66 ± 107.51
	ST	20.55 ± 0.35	20 ± 0.99	18.41 ± 1.59	15.33 ± 0.99
	BT	20.46 ± 0.45	19.32 ± 0.803	14.88 ± 2.11	6.99 ± 2.28
	SS	0.08 ± 0.02	9.96 ± 8.66	26.85 ± 4.48	33.45 ± 0.3
	BS	0.08 ± 0.02	12.7 ± 8.93	31.61 ± 1.45	33.82 ± 0.25
		Riverine	Estuarine	Shelf	Slope
Abundance	Z	6.91 ± 2.33	7.75 ± 2.06	28 ± 12.75	151.66 ± 107.51
	ST	20.4 ± 0.54	19.99 ± 1.21	18.41 ± 1.59	15.33 ± 0.99
	BT	20.28 ± 0.56	18.96 ± 0.75	14.88 ± 2.11	6.99 ± 2.28
	SS	1.15 ± 2.34	14.43 ± 8.98	26.85 ± 4.48	33.45 ± 0.3
	BS	1.2 ± 2.44	19.07 ± 5.04	31.61 ± 1.45	33.82 ± 0.25

Table 3. Estimates of mitochondrial DNA polymorphism in different fish models. Variables and phylogenetic informative sites in the total data set; haplotype diversity and nucleotide diversity (Nei 1987).

Species	<i>Brevoortia aurea</i>	<i>Engraulis anchoita</i>	<i>Squatina guggenheim</i>
mitochondrial <i>Cyt-b</i> Gene (bp)	720	720	448
Variable Sites	199	119	295
Phylogenetic informative Sites	88	60	128
Number of Haplotypes	46	14	15
Haplotype Diversity (SD)	1.000 (0.005)	0.978 (0.003)	0.382 (0.078)
Nucleotide diversity (SD)	0.06 (0.015)	0.060 (0.030)	0.011 (0.003)

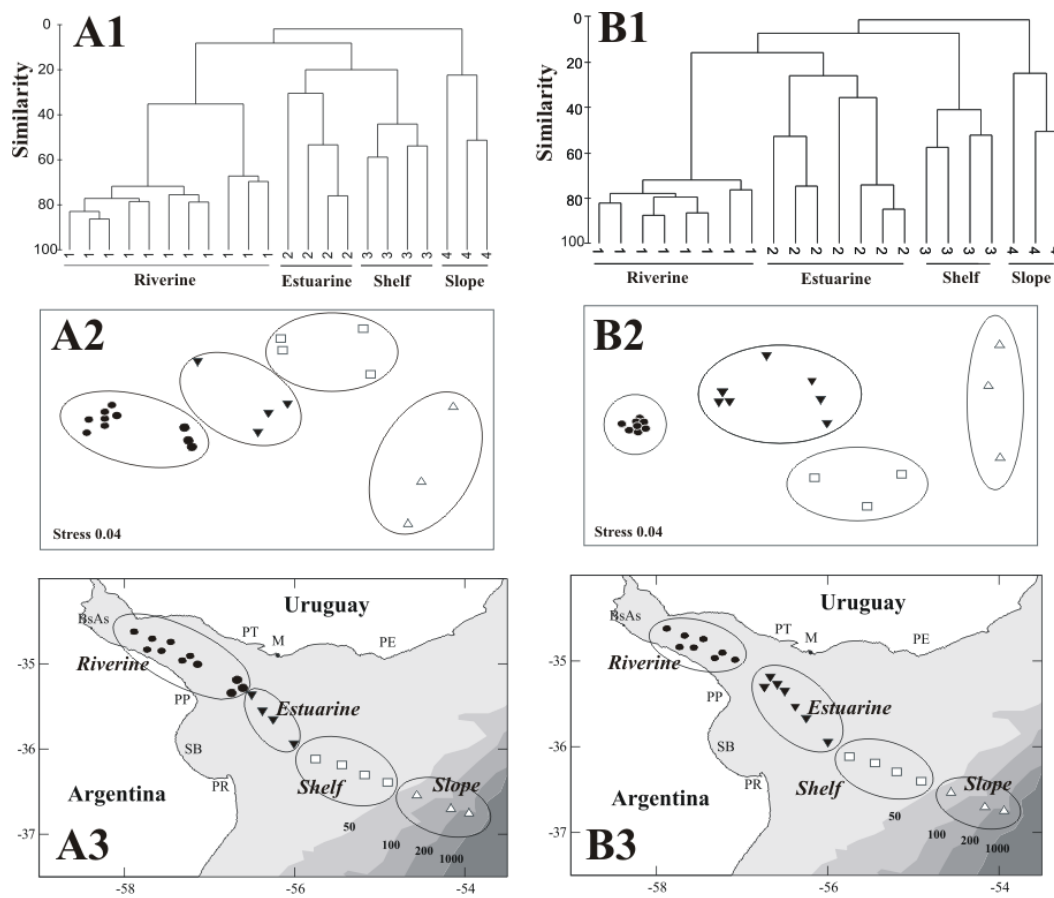


Figure 1. Dendrograms of the cluster analysis (1), nMDS diagrams (2) and location of the fish assemblage areas (3) defined for abundance (A) and biomass (B) analysis.

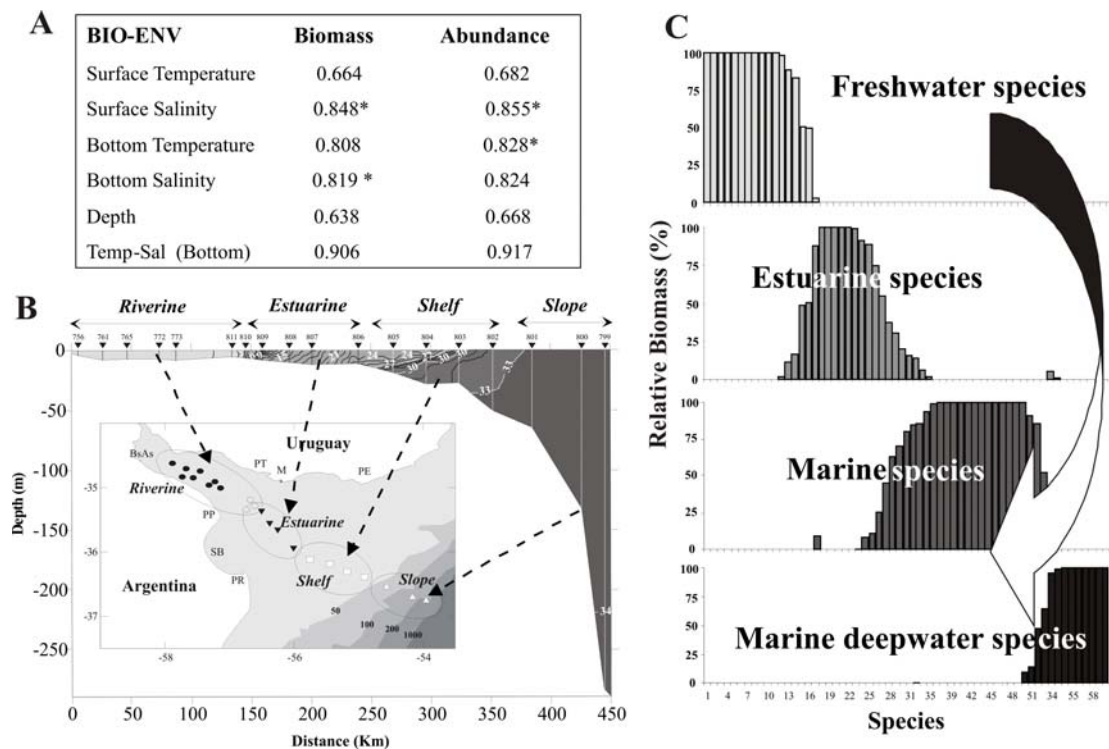


Figure. 2. A) Combination of the environmental variables yielding the best matches of biotic (species biomass or abundance, Bray Curtis similarity), and abiotic (Euclidean distance) similarity matrices, as measured by weighted Spearman correlation by means of BIO-ENV process. Z, depth (m); ST, surface temperature ($^{\circ}\text{C}$); BT, bottom temperature ($^{\circ}\text{C}$); SS, surface salinity; BS, bottom salinity. B) Location of the fish assemblage areas defined using biomass on the distribution of salinity along the sampling station, and their area of distribution. C) Relative biomass (%) of the species by fish assemblage area defined by multivariate analyses using biomass (t/nm^2). (1) *A. valenciennesi*, (2) *B. chauliodon*, (3) *C. carpio*, (4) *H. laplatae*, (5) *L. obtusidens*, (6) *P. bonariensis*, (7) *P. vetula*, (8) *P. maculatus*, (9) *P. lineatus*, (10) *P. granulosus*, (11) *N.barba*, (12) *R. dorbignyi*, (13)

P. albicans, (14) *L. pati*, (15) *P. valenciennesi*, (16) *L. grossidens*, (17) *M. furnieri*, (18) *A. marinii*, (19) *B. aurea*, (20) *C. orbignyana*, (21) *C. guatucupa*, (22) *P. brasiliensis*, (23) *M. ancylodon*, (24) *P. punctatus*, (25) *P. signata*, (26) *S. bonapartii*, (27) *P. patagonicus*, (28) *M. goodei*, (29) *T. lepturus*, (30) *M. schmitti*, (31) *R. agassizi*, (32) *S. guggenheim*, (33) *S. brasiliensis*, (34) *T. lathami*, (35) *A. brasilianus*, (36) *A. castelnaui*, (37) *D. auriga*, (38) *G. galeus*, (39) *M. argentinae*, (40) *Myliobatis* sp., (41) *P. pagrus*, (42) *P. brasiliensis*, (43) *P. nudigula*, (44) *P. semifasciata*, (45) *S. megalops*, (46) *T. lepidopodea*, (47) *Z. brevirostris*, (48) *D. tschudii*, (49) *X. rasile*, (50) *C. bergi*, (51) *M. hubbsi*, (52) *U. brasiliensis*, (53) *B. albesens*, (54) *C. c. marinii*, (55) *G. blacodes*, (56) *H. lahillei*, (57) *M. magallanicus*, (58) *P. ramsayi*, (59) *S. acanthias*, (60) *S. mitsukurii*.

Figure 3. Phylogenetic reconstruction using neighbour joining tree based on HKY85 model of molecular substitution. This model was obtained using hierarchical likelihood ratio tests (Huelsenbeck & Crandall, 1997) implemented in MODELTEST 3.06 (Posada & Crandall, 1998). Sixty individuals sampled belong to four collecting sites: Inner Shelf 35°S 53°W; Inner Shelf 36°S 54°W; Outer Shelf; Outer Río de la Plata. Numbers above branches represent 50% bootstrap supports.

Figure 4 Phylogenetic reconstruction using neighbour joining tree based on GTR model of molecular substitution. This model was obtained using hierarchical likelihood ratio tests (Huelsenbeck & Crandall, 1997) implemented in MODELTEST 3.06 (Posada & Crandall, 1998). Seventy individuals sampled belong to three collecting sites: Atlantic Ocean coast; Outer Shelf; Outer Río de la Plata. Numbers above branches represent 50% bootstrap supports.